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Mahesh K. Lakshman^{ab}; Roland E. Lehr^a

^a Department of Chemistry, University of Oklahoma, Norman, OK ^b Section on Oxidation Mechanisms, Laboratory of Bioorganic Chemistry, NIDDK, The National Institutes of Health, Bethesda, MD

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SOLVENT DEPENDENT CHANGES IN THE PROTON NMR SPECTRA OF 2'-DEOXYADENOSINE AND ITS DERIVATIVES

Mahesh K. Lakshman[†] and Roland E. Lehr* Department of Chemistry, University of Oklahoma, Norman OK 73019.

Abstract: Intramolecular hydrogen bonding in four purine-2'-deoxyribosides containing free sugar hydroxyls has been observed in CDCl₃. This bond results in the appearance of a *double doublet* for H-1' of the C-6 substituted β -anomers. On increasing the solvent polarity, as in DMSO-d_{θ} the signal appears as triplet, which is typical of previously reported NMR spectra of β -anomers in D₂O and DMSO-d_{θ}. Solvent polarity, therefore plays a significant role in determining the splitting patterns of the anomeric proton in these compounds.

Intramolecular hydrogen bonding between the C-5' hydroxyl group and N-3 of the purine has been reported for purine deoxyribosides in the solid state, and numerous X-ray crystal structures indicating these have been reported in the literature.\(^1\) In the solution state also, such hydrogen bonding has been reported, but for purine ribosides.\(^2\),\(^3\) This is due to the fact that purine nucleosides can be appropriately derivatized in order to increase their otherwise limited solubility in relatively non-polar solvents. In two such typical examples, the 2',3'-acetonide was synthesized.\(^3\),\(^4\) In these cases, both the conformational freedom of the furanose ring and the number of hydrogen bonding sites are reduced. Further, the changes observed in the anomeric H-1' resonance of the purine ribosides on changing solvent polarity are limited to changes in the doublet for this proton.

To our knowledge, ¹H NMR data in non-polar solvents for purine-2'-deoxyribosides with free sugar hydroxyl groups have not been reported. Interestingly, the H-1' of these compounds show remarkably different splitting patterns in solvents of differing polarities. The α - and β - anomers of purine-2'-deoxyribosides have commonly been distinguished based upon the appearance of a double doublet or a triplet, respectively for

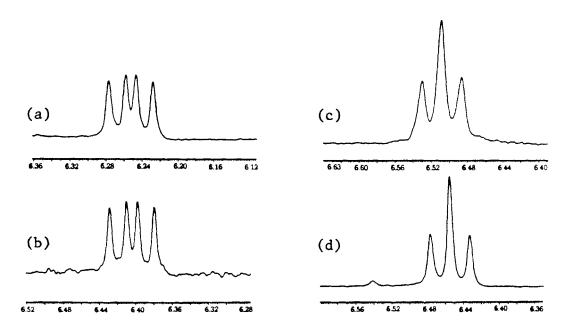
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H-1'.5,6,7,8 We report here that H-1' of C-6 substituted β -anomers also appears as a *double doublet* when the NMR solvent is of relatively low polarity (CDCl₃) and intramolecular hydrogen bonding between the C-5' hydroxyl group and N-3 of the purine occurs.

In all of the cases examined by NMR spectroscopy in the literature, the solvents used in determining the anomeric configuration were either D₂O or DMSO-d₆. In our study, we found that the purine deoxyribosides are sufficiently soluble in CDCl₃ to obtain fourier transformed ¹H NMR spectra. Spectra (Figure 2) for compounds 1 to 5 (Figure 1) were obtained in both CDCl₃ and DMSO-d₆.⁵ In the latter solvent, compounds 1 to 4 exhibit the expected $triplet, ^{6,7,8,9}$ whereas in $CDCl_3$ they exhibit a $double\ doublet$ for H-In addition, as is explained later, the C-5' protons show a splitting pattern that is consistent with the 5'-OH to N-3 hydrogen bond, in CDCl₂. In the absence of any second order couplings, the observed changes in the multiplicity of H-1' can be attributed solely to changes in the sugar ring conformation. Such changes, if any, would also be reflected in the splitting patterns of the other sugar ring protons. In CDCl₃ each H-2' appears as a septet and a double doublet, respectively, whereas in DMSO-d₆, these protons appear as a quintet and a doublet of doublet of doublets. In addition, in CDCl3 H-4' appears as a singlet and H-3' appears as a doublet and the dihedral angle of one H-2' to H-3' is nearly 90°. On using DMSO-dgo H-4' and H-3' appear as a *quartet* and a broadened signal, respectively. The effects observed on changing solvent polarity are consistent with a change in the sugar conformation, the cause of which is the formation of an intramolecular hydrogen bond between the 5'-OH and N-3. Such a hydrogen bond is also expected to affect each H-5' resonance. explained later this is indeed the case. In CDCl₃, in order to establish the intramolecular hydrogen bond a syn-conformation at C-1' and a 2'-endo sugar pucker are essential for close proximity of the hydrogen bonding atoms. Under these circumstances, H-1' has different dihedral angles to each H-2'. In DMSO-d₆, on the other hand, the hydroxyl protons can hydrogen bond to solvent, resulting in a predominance of the 3'-endo conformation, wherein the dihedral angles of H-1' with each H-2' are nearly equal resulting in the observed splitting patterns (vide supra).

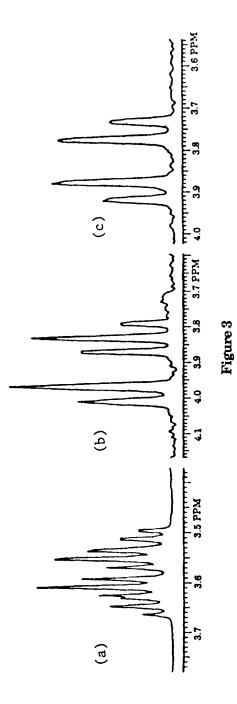
Due to the lower solubility of 1 and 2 in CDCl₃ small amounts of MeOH- d_4 were added to enable the spectra to be observed (50 μ L for 1 and 90 μ L for 2 in 0.8 mL CDCl₃). Under these conditions 1 [Figure 2(a)] and 2 (data not shown) exhibit a *double doublet* for H-1'. The effect of solvent on the H-1' multiplicity of 3 is shown in Figures 2(b), (c) and (d). In CDCl₃, H-1'

Figure 1



(a) H-1' of 1 in CDCl₃; H1' of 3 in (b) CDCl₃ (c) MeOH-d₄ (d) DMSO-d₆

Figure 2



Diastereotopic H-5' of $\underline{3}$ in : (a) DMSO- d_6 (b) CDCl $_3$ (c) CDCl $_3$ +MeOH- d_4

appears as a double doublet, in MeOH-d₄ the central signals approach coalescence and in DMSO-d₆ the signal is a distinct triplet.

Figure 3(a) shows the multiplicity of the C-5' protons of 3 in DMSO-de whereas Figures 3(b) and (c) show the multiplicities in CDCl₃ and CDCl₃ with a few drops of MeOH-d4 added, respectively. In CDCl3 the coupling of each H-5' with H-4' is very small (H-4' appears as a singlet at δ 4.22 ppm) and the individual C-5' protons appear as a doublet and a triplet, respectively. In a rigid framework established by the 5'-OH to N-3 hydrogen bond, one H-5' has a large dihedral angle with the hydroxyl proton (~180°, J = 10.8 Hz), whereas the other has a smaller angle (~ 40° , and in the case of 4 the smaller H-5' to OH coupling is observed, J = 2.4 Hz). Addition of a few drops of MeOH-d4 results in loss of the hydroxyl proton coupling and the resulting signals are a pair of doublets ($J_{gem} = 12.7 \text{ Hz}$). Also consistent with this hydrogen bond, in $CDCl_3$ the 5'-OH appears downfield (δ 5.0 ppm) as a doublet ($J_{OH, H5'} = 10.8 \text{ Hz}$), while the 3'-OH appears at δ 1.9 ppm. In DMSO-d_e, on the other hand, the 5'-OH appears as a triplet, indicating its coupling with both C-5' protons. Further, both the 3'- and the 5'-OH appear downfield (δ 5.4 and 4.88 ppm, respectively)¹⁰, presumably due to hydrogen bonding of both hydroxyl protons to the solvent. Compound 4 shows identical spectral features as 3 in both CDCl3 and DMSO-d6. Table 1 shows the chemical shifts and coupling constants of H-1' in 1 to 6.

Robins and Robins, in their initial paper describing anomer determination by ¹H NMR, have mentioned that substitutents on the heterocycle or the sugar could alter the sugar pucker. ^{6,9} This indeed is the case. Compounds 5 and 6 show a triplet for H-1'. Acetylation or silylation of the hydroxyl groups in 4 and silylation of the hydroxyl groups in 2 also results in a triplet for H-1' in CDCl₃, whereas the parent compounds exhibit a double doublet. ¹⁰ It is quite possible that inhibition of the intramolecular hydrogen bonding results in a 3'-endo conformation in these compounds.

Thus, in a solvent such as chloroform with a low dielectric constant (ϵ = 5.62), intramolecular hydrogen bonding causes a predominance of the 2'-endo conformation, resulting in the appearance of a double doublet for H-1'. On increasing the dielectric constant to that of DMSO (ϵ = 46.68), intermolecular hydrogen bonding apparently becomes preferred, and a triplet is observed for H-1' in DMSO-d₆. In addition to a variety of factors such as anomeric configuration, substitution on the sugar hydroxyls and the purine, attention must be paid to another variable, namely solvent polarity, when using the H-1' splitting patterns for anomer assignment in C-6 substituted purine-2'-deoxyribosides.

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COMPOUND	SOLVENT	MULTIPLICITY	SHIFT (δ ppm)	J (Hz)
1	DMSO-d ₆	triplet	6.34	7.8
	CDCl ₃	double doublet	6.25	9.2, 5.5
2	DMSO-d ₆	triplet	6.40	6.6
	CDCl ₃	double doublet	6.26	8.3, 5.8
3	DMSO-de	triplet	6.46	6.5
	CDCl ₃	double doublet	6.40	9.3, 5.6
4	DMSO-d ₆	triplet	6.48	6.5
	CDCl ₃	double doublet	6.41	9.5, 5.5
5	DMSO-d ₆	triplet	6.41	6.6
_	CDCl ₃	triplet	6.42	7.1
6	CDC13	triplet	6.40	6.8

Table 1: Chemical shifts and coupling constants for H-1' in 1-6.*

Note: Compounds 1-6 were prepared through published procedures. 12,13 Compound 1: 2 mg were dissolved in 0.8 mL CDCl₃ and 50 μ L MeOH-d₄, 128 scans were acquired with a line broadening of 0.5.

Compounds 3 and 4: 1 mg of each compound was dissolved in 0.8 mL CDCl₃, 128 scans were acquired with a line broadening of 0.5.

Compound 2: A saturated solution was prepared in 0.8 mL $CDCl_3$ and 90 μL MeOH- d_4 . Usually a large number of scans are needed for a good spectrum with a line broadening of 0.5.

Compounds 1 to 4:2 mg of each was dissolved in 0.8 mL DMSO-d_{θ} 128 scans were acquired with a line broadening of 0.5.

No resolution enhancement was used while acquiring data for these compounds and all spectra were obtained at room temperature.

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- † Present address: Section on Oxidation Mechanisms, Laboratory of Bioorganic Chemistry, NIDDK, The National Institutes of Health, Bethesda, MD 20892.
- This work was supported in part, by grant CA 22985 from the National Cancer Institute, DHHS, to R.E.L.
 For some typical examples see:

^{*}Spectra were recorded at 300 MHz.

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- 5. As typical examples the entire 1 H NMR spectra of 3 and 4 are: 3 in DMSO-d₆: 2.36 (ddd, 1 H_{2'a}, 1 J = 13.4, 6.3, 4.0 Hz); 2.75 (quint, 1 H_{2'b}, 1 J = 6.3 Hz); 3.50-3.70 (m, 2 H_{5'}); 3.88 (q, 1 H_{4'}, 1 J = 4.3 Hz); 4.44 (br, 1 H_{3'}); 4.88 (t, 1 OH_{5'}, 1 J = 5.3 Hz); 5.40 (d, 1 OH_{3'}, 1 J = 4.0 Hz); 6.46 (t, 1 H_{1'}, 1 J = 6.6 Hz); 8.79 (s, 1 H₂); 8.88 (s, 1 H₉).
 - 4 in DMSO-d₆: 2.36 (ddd, $1\rm{H}_{2'a}$, J = 13.4, 6.2, 4.0 Hz); 2.75 (quint, $1\rm{H}_{2'b}$, J = 6.2 Hz); 3.50-3.70 (m, $2\rm{H}_{5'}$); 3.88 (q, $1\rm{H}_{4'}$, J = 4.4 Hz); 4.44 (br, $1\rm{H}_{3'}$); 4.88 (t, $O\rm{H}_{5'}$, J = 5.4 Hz); 5.40 (d, $O\rm{H}_{3'}$, J = 4.0 Hz); 6.48 (t, $1\rm{H}_{1'}$, J = 6.7 Hz); 8.79 (s, $1\rm{H}_{2}$); 8.88 (s, $1\rm{H}_{8}$).
 - 3 in CDCl₃: 1.90 (d, O H_3 ', J = 3.2); 2.39 (dd, 1 $H_{2'a}$, J = 13.5, 5.5 Hz); 3.04 (septet, 1 $H_{2'b}$, J = 5.3 Hz); 3.70 (t, 1 $H_{5'a}$, J = 10.8 Hz); 3.95 (d, 1 $H_{5'b}$, J = 12.7 Hz); 4.22 (s, 1 $H_{4'}$); 4.81 (d, 1 H_3 ', J = 4.8 Hz); 5.00 (d, O H_5 ', J = 10.8 Hz); 6.39 (dd, 1 $H_{1'}$, J = 9.3, 5.6 Hz); 8.21 (s, 1 H_2); 8.74 (s, 1 H_8).
 - 4 in CDCl₃: 1.90 (d, O H_3 ', J = 3.2); 2.40 (dd, 1 $H_{2'a}$, J = 13.4, 5.5 Hz); 3.04 (septet, 1 $H_{2'b}$, J = 5.3 Hz); 3.78 (t, 1 $H_{5'a}$, J = 11.1 Hz); 3.95 (dd, 1 $H_{5'b}$, J = 12.6, 2.4 Hz); 4.22 (s, 1 $H_{4'}$); 4.81 (br, 1 $H_{3'}$); 5.10 (dd, O $H_{5'}$, J = 10.9, 2.4 Hz); 6.40 (dd, 1 $H_{1'}$, J = 9.5, 5.5 Hz); 8.18 (s, 1 H_2); 8.63 (s, 1 H_8).
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10. Chemical shift (coupling constant) in DMSO-d₆ for:

	5'-O <u>H</u>	3'-O H
1	5.23 (6.4 Hz)	5.29 (3.6 Hz)
2	5.00 (5.7 Hz)	5.30 (2.2 Hz)
3	4.88 (5.3 Hz)	5.40 (4.0 Hz)
4	4.88 (5.7 Hz)	5.40 (4.0 Hz)

11. H-1' in the:

3',5'-diacetate derivative of 4 is at δ 6.50 ppm; $J_{1',2'}$ = 6.3 Hz

3',5'-di- t BuMe $_{2}$ Si derivative of 4 is at δ 6.51 ppm; $J_{1'.2'}$ = 6.4 Hz

3',5'-di- $^{\rm t}$ BuMe $_{\rm 2}$ Si derivative of 2 is at δ 6.40 ppm; $J_{1',2'}$ = 6.4 Hz

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